



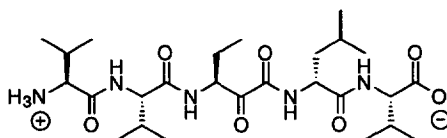
## A Convergent Synthesis of Poststatin: Application of the Acyl Cyanophosphorane Coupling Procedure in the Formation of a Peptidic $\alpha$ -Keto Amide.

Harry H. Wasserman\* and Anders K. Petersen

Department of Chemistry, Yale University, New Haven, CT 06520-8107, U.S.A.

**Abstract:** A convergent synthesis of the pentapeptide poststatin has been developed. The key step involves oxidative cleavage of an acyl cyanophosphorane. The resulting  $\alpha,\beta$ -diketo nitrile is then coupled to the free amine of a C-terminal-dipeptidyl component to generate the protected natural product. Deprotection by hydrogenolysis furnishes poststatin. Copyright © 1997 Elsevier Science Ltd. © 1997, Elsevier Science Ltd. All rights reserved.

Poststatin<sup>1</sup> (**1**), an inhibitor of prolyl endopeptidase, is a naturally occurring pentapeptide isolated from *Streptomyces viridochromogenes*. The amino acid sequence is: H-Val-Val-Pos-D-Leu-Val-OH, where Pos is the abbreviation for the unusual (*S*)-3-amino-2-oxopentanoic acid named L-postine. The  $\alpha$ -keto-amide group, which is present in a number of cyclic peptides,<sup>2</sup> appears to be necessary for the biological activity of poststatin.<sup>1b</sup> The cyclotheonamides<sup>3</sup> contain the  $\alpha$ -keto homologue of arginine, whereas the structurally related orbiculamides,<sup>4</sup> keramamides,<sup>5</sup> and discobahamins<sup>6</sup> incorporate leucine extended with an  $\alpha$ -carbonyl group. In the eurystatins,<sup>7</sup> (*S*)-3-amino-2-oxobutanoic acid is believed to be responsible for the prolyl endopeptidase inhibitory effect, while the tricarbonyl amino acid 4-amino-2,3-dioxo-6-methylheptanoic acid comprises the electrophilic binding site in the depsipeptides YM-47141 and YM-47142.<sup>8</sup> Examples of peptide alkaloids derived from  $\alpha$ -keto amides include verongamine,<sup>9</sup> eusynstyelamide,<sup>10</sup> and the anchinopeptolides.<sup>11</sup> Other studies have shown, that rationally designed derivatives of  $\alpha$ -keto acids represent an important class of enzyme inhibitors.<sup>12</sup>



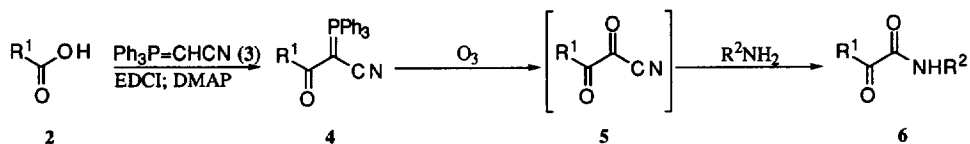
**1** Poststatin: H-Val-Val-Pos-D-Leu-Val-OH; Pos = Postine

In a previous synthesis of poststatin,<sup>1d,f</sup> a linear approach employed (*2R,3S*)-3-amino-2-hydroxypentanoic acid as a precursor of postine. This procedure followed traditional lines, whereby 2-keto carboxylates are prepared through oxidation of the intermediate hydroxy compounds, which may be obtained

from the appropriate 2-hydroxy trithioorthoformate,<sup>3b,13</sup> cyanohydrin,<sup>14</sup> 2-(hydroxymethyl)furan,<sup>15</sup> or other 2-hydroxy carboxylate precursors.<sup>12,16,17</sup> In general, these syntheses have disadvantages associated with many steps or racemization.<sup>18</sup>

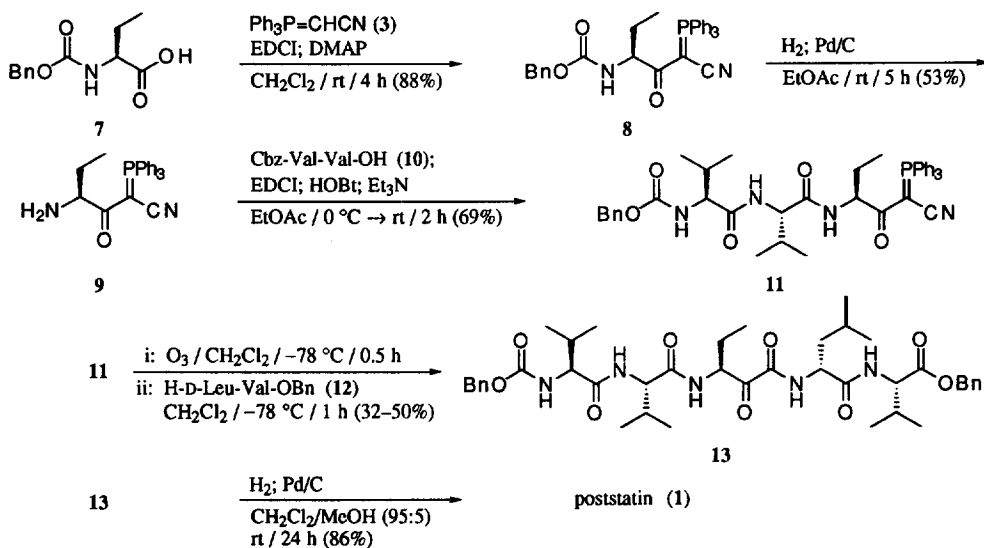
As part of a research program focused on the synthesis of peptidyl enzyme inhibitors,<sup>19a</sup> we have developed a novel method for the efficient construction of  $\alpha$ -keto amides by acylation of amines with  $\alpha,\beta$ -diketo nitriles (cf. Scheme I).<sup>19b</sup> This procedure has now been applied in a total synthesis of poststatin.

**Scheme I. Preparation of  $\alpha$ -Keto Amides.**



In a retrosynthetic sense, poststatin may be regarded as postine substituted with a dipeptide in each direction. Two ways were considered for assembling these fragments. In one route, the *N*-terminal may be initially coupled to *N*-protected valylvaline, followed by *C*-terminal chain elongation with *O*-protected D-leucylvaline. Alternatively, the order of the peptide couplings may be reversed. We chose the former strategy, because it has the advantage of generating the  $\alpha$ -keto-amide functionality at a late stage in the synthesis, thereby minimizing epimerization of the postine residue. As outlined in Scheme II, the key bond-forming step is the condensation of Cbz-Val-Val-Pos-CN and H-D-Leu-Val-OBn (**12**). The activated acylating agent is generated in situ by ozonolysis of the corresponding acyl cyanophosphorane **11**. Thus, the unmasking of the  $\alpha$ -keto-carboxylate and the peptide coupling reaction take place in one synthetic operation.

**Scheme II. Synthesis of Poststatin.<sup>20</sup>**



The acyl cyanophosphorane **8** was formed (88%) by the coupling of commercially available Cbz-protected (S)-(+)-2-aminobutanoic acid (**7**) with (cyanomethylene)triphenylphosphorane (**3**)<sup>21,22</sup> using EDCI and DMAP. Removal of the protecting group was accomplished by hydrogenolysis over Pd/C.<sup>23</sup> The crude amine was coupled with Cbz-protected valylvaline<sup>24</sup> under standard peptide-coupling conditions to yield the crystalline tripeptide **11** (69%) as the sole epimer. Oxidative cleavage of the carbon-phosphorus double bond, by reaction with ozone, generated the highly electrophilic  $\alpha,\beta$ -diketo nitrile intermediate corresponding to **5**. Immediate addition of the free amine<sup>25</sup> of D-leucylvaline *O*-benzyl ester (**12**)<sup>1d,f</sup> produced the protected pentapeptide (**13**).<sup>26</sup> Essentially no epimerization of the poststine unit was observed at this stage. Hydrogenolysis of the benzylic protecting groups furnished poststatin (**1**)<sup>27</sup> (86%). NMR examination of the final product suggested the presence of ca 15% of a poststatin epimer.<sup>1d,f</sup>

The possibility of establishing the  $\alpha$ -keto-amide linkage between simple substrates followed by bidirectional synthetic manipulations is currently being investigated. If successful, this versatile methodology may be applied to peptides such as the orbiculamides,<sup>4</sup> keramamides,<sup>5</sup> discobahamins,<sup>6</sup> and other target molecules containing substructural elements which are sensitive to the oxidative conditions.

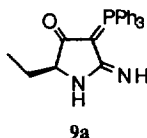
### Acknowledgments

Support of this work by Grants from the National Institutes of Health and the National Science Foundation is gratefully acknowledged. We thank Dr. Y. Muraoka of the Institute of Microbial Chemistry, Tokyo, for a sample of synthetic poststatin.

### References and Notes

- (a) Aoyagi, T.; Nagai, M.; Ogawa, K.; Kojima, F.; Okada, M.; Ikeda, T.; Hamada, M.; Takeuchi, T. *J. Antibiot.*, **1991**, *44*, 949–955. (b) Nagai, M.; Ogawa, K.; Muraoka, Y.; Naganawa, H.; Aoyagi, T.; Takeuchi, T. *J. Antibiot.*, **1991**, *44*, 956–961. (c) Tsuda, M.; Muraoka, Y.; Nagai, M.; Aoyagi, T.; Takeuchi, T. *Peptide Chemistry*, **1991**, 223–228. (d) Takeuchi, T.; Aoyagi, T.; Hamada, M.; Naganawa, H.; Ogawa, K.; Nagai, M.; Muraoka, Y.; Tsuda, M. U.S. Patent 5,359,138, **1994**. (e) Tsuda, M.; Muraoka, Y.; Nagai, M.; Aoyagi, T.; Takeuchi, T. *J. Antibiot.*, **1996**, *49*, 281–286. (f) Tsuda, M.; Muraoka, Y.; Nagai, M.; Takeuchi, T.; Aoyagi, T. *J. Antibiot.*, **1996**, *49*, 287–291. (g) Tsuda, M.; Muraoka, Y.; Nagai, M.; Aoyagi, T.; Takeuchi, T. *J. Antibiot.*, **1996**, *49*, 890–899. (h) Tsuda, M.; Muraoka, Y.; Someno, T.; Nagai, M.; Aoyagi, T.; Takeuchi, T. *J. Antibiot.*, **1996**, *49*, 900–908. (i) Tsuda, M.; Muraoka, Y.; Nagai, M.; Aoyagi, T.; Takeuchi, T. *J. Antibiot.*, **1996**, *49*, 909–920.
- Fusetani, N.; Matsunaga, S. *Chem. Rev.* **1993**, *93*, 1793–1806.
- (a) Fusetani, N.; Matsunaga, S.; Matsumoto, H.; Takebayashi, Y. *J. Am. Chem. Soc.* **1990**, *112*, 7053–7054. (b) Hagihara, M.; Schreiber, S. L. *J. Am. Chem. Soc.* **1992**, *114*, 6570–6571.
- Fusetani, N.; Sugawara, T.; Matsunaga, S. *J. Am. Chem. Soc.* **1991**, *113*, 7811–7812.
- Kobayashi, J.; Itagaki, F.; Shigemori, H.; Ishibashi, M.; Takahashi, K.; Ogura, M.; Nagasawa, S.; Nakamura, T.; Hirota, H.; Ohta, T.; Nozoe, S. *J. Am. Chem. Soc.* **1991**, *113*, 7812–7813.
- Gunasekera, S. P.; Pomponi, S. A.; McCarthy, P. J. *J. Nat. Prod.* **1994**, *57*, 79–83.
- Toda, S.; Kotake, C.; Tsuno, T.; Narita, Y.; Yamasaki, T.; Konishi, M. *J. Antibiot.*, **1992**, *45*, 1580–1586.
- Orita, M.; Yasumuro, K.; Kokubo, K.; Shimizu, M.; Abe, K.; Tokunaga, T.; Kaniwa, H. *J. Antibiot.*, **1995**, *48*, 1430–1434.
- Mierzwa, R.; King, A.; Conover, M. A.; Tozzi, S.; Puar, M. S.; Patel, M.; Coval, S. J.; Pomponi, S. A. *J. Nat. Prod.* **1994**, *57*, 175–177.
- Swersey, J. C.; Ireland, C. M.; Cornell, L. M.; Peterson, R. W. *J. Nat. Prod.* **1994**, *57*, 842–845.
- Casapullo, A.; Minale, L.; Zollo, F.; Lavayre, J. *J. Nat. Prod.* **1994**, *57*, 1227–1233.
- (a) Burkhart, J. P.; Peet, N. P.; Bey, P. *Tetrahedron Lett.* **1988**, *29*, 3433–3436. (b) Angelastro, M. R.; Peet, N. P.; Bey, P. *J. Org. Chem.* **1989**, *54*, 3913–3916. (c) Burkhart, J. P.; Peet, N. P.; Bey, P.

- Tetrahedron Lett.* **1990**, *31*, 1385–1388. (d) Peet, N. P.; Burkhart, J. P.; Angelastro, M. R.; Giroux, E. L.; Mehdi, S.; Bey, P.; Kolb, M.; Neises, B.; Schirlin, D. *J. Med. Chem.* **1990**, *33*, 394–407. (e) Iwanowicz, E. J.; Lin, J.; Roberts, D. G. M.; Michel, I. M.; Seiler, S. M. *BioMed. Chem. Lett.* **1992**, *2*, 1607–1612. (f) Edwards, P. D.; Meyer, E. F., Jr.; Vijayalakshmi, J.; Tuthill, P. A.; Andisik, D. A.; Gomes, B.; Strimpler, A. *J. Am. Chem. Soc.* **1992**, *114*, 1854–1863. (g) Cutrona, K. J.; Sanderson, P. E. *J. Tetrahedron Lett.* **1996**, *37*, 5045–5048.
13. Beautement, K.; Clough, J. M. *Tetrahedron Lett.* **1987**, *28*, 475–478.
  14. Wipf, P.; Kim, H.-Y. *Tetrahedron Lett.* **1992**, *33*, 4275–4278.
  15. Deng, J.; Hamada, Y.; Shioiri, T.; Matsunaga, S.; Fusetani, N. *Angew. Chem., Int. Ed. Engl.* **1994**, *33*, 1729–1731.
  16. Schmidt, U.; Weinbrenner, S. *J. Chem. Soc., Chem. Commun.* **1994**, 1003–1004.
  17. Sowinski, J. A.; Toogood, P. L. *Tetrahedron Lett.* **1995**, *36*, 67–70.
  18. In the Dakin–West reaction, for example, a 3-*N*-acylamino-2-keto ester is formed directly from a 2-*N*-acylamino acid. However, the product is completely racemized due to the intermediacy of an enamide. Charles, I.; Latham, D. W. S.; Hartley, D.; Oxford, A. W.; Scopes, D. I. C. *J. Chem. Soc., Perkin Trans. 1* **1980**, 1139–1146.
  19. (a) Wasserman, H. H.; Ennis, D. S.; Power, P. L.; Ross, M. J.; Gomes, B. *J. Org. Chem.* **1993**, *58*, 4785–4787. (b) Wasserman, H. H.; Ho, W.-B. *J. Org. Chem.* **1994**, *59*, 4364–4366.
  20. New compounds were characterized by TLC, mp (for solids), opt. rot., IR, <sup>1</sup>H NMR, <sup>13</sup>C NMR, MS, and HRMS.
  21. (a) Trippett, S.; Walker, D. M. *J. Chem. Soc.* **1959**, 3874–3876. (b) Schiemenz, G. P.; Engelhard, H. *Chem. Ber.* **1961**, *94*, 578–585. (c) Bestmann, H. J.; Pfohl, S. *Liebigs Ann. Chem.* **1974**, 1688–1693.
  22. Improved procedure for the preparation of (cyanomethylene)triphenylphosphorane 3:<sup>21</sup> (Cyanomethyl)triphenylphosphonium chloride (Lancaster Synthesis Inc.) (123.6 g, 0.37 mol) was suspended in dry dichloromethane (1.8 L). Triethylamine (127 mL, 92.2 g, 0.91 mol) was added over a period of 15 min. The clear yellow solution was stirred for a further 25 min, before it was washed with prechilled water (2 × 500 mL) and dried with magnesium sulfate (1 h). Filtration and concentration afforded a pale yellow solid (108.1 g, 0.36 mol, 98% yield). The crude ylide was contaminated with less than 5% triphenylphosphine oxide and could be condensed successfully with carboxylic acids. The product was purified by recrystallization from benzene (1.5 L, 14 mL/g) to give an off-white product (87.0 g, 0.29 mol, 79%).
  23. Along with amine **9** (53%), the cyclic amidine **9a** (21%) formed by intramolecular aminolysis of the cyano group (26% unconverted **8**).



24. Prepared from valylvaline (85%) by reaction with benzyl chloroformate.
25. The dipeptidyl amine **12** was obtained (85%) from the hydrotrifluoroacetate of D-leucylvaline *O*-benzyl ester<sup>1d,f</sup> by treatment with saturated aqueous sodium hydrogen carbonate.
26. When conducted on a 5.5 mmol scale, a 32% yield of the pentapeptide **13**<sup>1d,f</sup> was obtained in impure state after chromatography. At 0.1 mmol scale the yield was consistently in the 50% range, and the product quality was significantly higher.
27. The crude poststatin was purified by reversed-phase-silica-gel flash chromatography using methanol/water = 60:40 as the eluent. The synthetic product was identical in all respects to a sample of synthetic poststatin, kindly provided by Dr. Muraoka. Due to the pronounced concentration dependence of the <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra, the final comparison was achieved by mixing equal amounts of the two samples. Under these circumstances, a single set of signals was observed.

(Received in USA 25 November 1996; revised 12 December 1996; accepted 16 December 1996)